MASSON TRICHROME (for FFPE)
Rowley
(last update 9/2021)

Date: ___/___/___ Technician (Initials): ______________

OBJECTIVE: Distinguishes collagen from muscle.

RECOMMENDED CONTROL TISSUE: Appendix, Small Intestine, Uterus

ID OF CONTROL USED: ________________________ Type of Tissue: ______________________________

<table>
<thead>
<tr>
<th>Reagent</th>
<th>Vendor</th>
<th>Catalog #</th>
<th>Was this a new Bottle (Y/N)? If Yes, check lot log.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bouin’s</td>
<td>Rowley</td>
<td>SO-306</td>
<td></td>
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<tr>
<td>Weigert’s Iron Hematoxylin A</td>
<td>Rowley</td>
<td>SO-465</td>
<td></td>
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<tr>
<td>Weigert’s Iron Hematoxylin B</td>
<td>Rowley</td>
<td>SO-466</td>
<td></td>
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<tr>
<td>Biebrich Scarlet-Acid Fuchsin</td>
<td>Rowley</td>
<td>F367-3</td>
<td></td>
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<tr>
<td>Phosphotungstic/Phosphomolybdic Acid</td>
<td>Rowley</td>
<td>F367-4</td>
<td></td>
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<tr>
<td>Aniline Blue</td>
<td>Rowley</td>
<td>F367-5</td>
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</tbody>
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REAGENT PREPARATION:
Working Weigert’s Iron Hematoxylin at 1:1 ratio of A:B

PROTOCOL

1. FOR FFPE SLIDES: Deparaffinize slides and bring to dH2O
2. Mordant tissue in Bouin’s solution @65°C 30 minutes.
3. Cool sections; Wash in running tap H2O until all yellow color is removed for 5 minutes.
4. Stain nuclei with Working (1A:1B) Weigert’s iron Hematoxylin for 15 minutes
5. Wash in running Tap H2O for 5 minutes
6. Rinse with dH2O
7. Stain in Biebrich Scarlet- Acid Fuchsin for 15 minutes
8. Rinse with DH2O
9. Place sections in working Phosphotungstic / Phosphomolybdic Acid solution for 10 minutes. Do not rinse. (discard used solution)
10. Stain sections with Aniline Blue solution for 15 minutes
11. Rinse in DH2O
12. QUICKLY Dehydrate in 95% and 100% ETOH
13. Clear in Xylene and mount

EXPECTED RESULTS: Nuclei - black; Cytoplasm, Keratin, Muscle fibers – red; Collagen, Mucin - blue

CASE IDS/SLIDE NUMBERS: