**PAS**  
(last update 09/2021)

Date: ___/___/___  Technician (Initials): ______________

**OBJECTIVE:** Staining method for carbohydrates.  
**RECOMMENDED CONTROL TISSUE:** Liver/Kidney

ID of Control Used: _________________________  Type of Tissue: _________________________

<table>
<thead>
<tr>
<th>Reagent</th>
<th>Vendor</th>
<th>Catalog #</th>
<th>Was this a new Bottle (Y/N)? If Yes, check lot log.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Schiff Reagent</td>
<td>Sigma</td>
<td>395-B</td>
<td></td>
</tr>
<tr>
<td>0.5% Periodic Acid</td>
<td>Fisher Scientific</td>
<td>BP581-100</td>
<td></td>
</tr>
<tr>
<td>Mayers Hematoxylin</td>
<td>Rowley</td>
<td>L-756-1A</td>
<td></td>
</tr>
</tbody>
</table>

**REAGENT PREPARATION (if needed):**

0.5% Periodic Acid Solution: 2.5 g in 500 ml MilliQ H2O  
*Reagent is stable for 1 year at room temperature.

**PROTOCOL**

NOTE: Allow Schiff Reagent to reach room temperature before staining.

1. Deparaffinize slides and bring to dI H2O  
2. Soak slides in 0.5% Periodic Acid 5 minutes  
3. Rinse in DH2O  
4. Soak slides in Schiff Reagent 20 minutes  
5. Wash in running Tap H2O 5 minutes  
6. Counterstain with Mayers Hematoxylin 2 minutes  
7. Rinse with Tap H2O  
8. Blue with Scott’s Tap H2O 1 minute  
9. Rinse with dI H2O  
10. Dehydrate clear and mount

**EXPECTED RESULTS:** Carbohydrates and Biological structures - Pink to Magenta; Nuclei- Blue

**CASE ID/s/SLIDE NUMBERS:**